3DOR 2022, 1-2 September, Florence, Italy

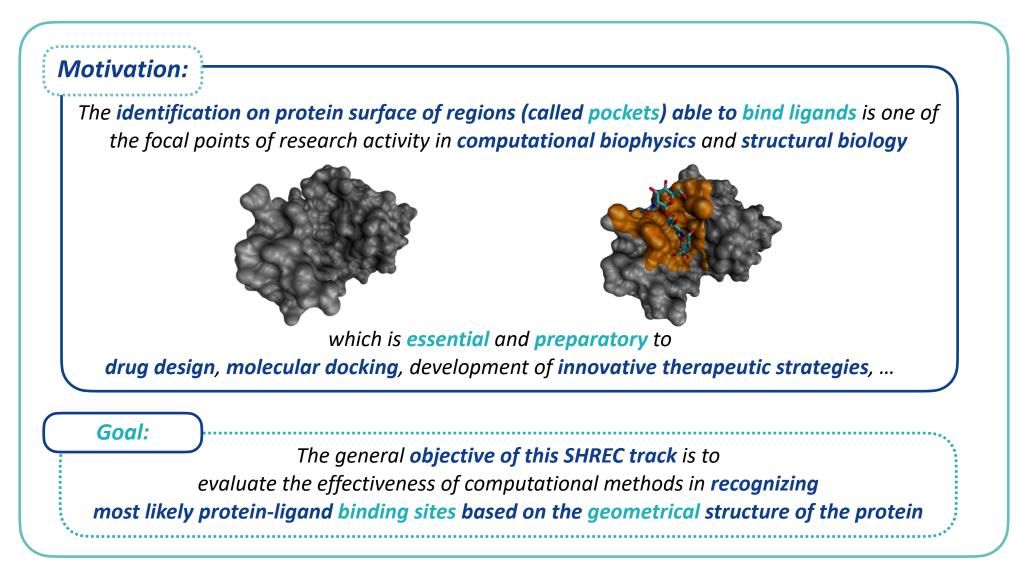
SHREC 2022: Protein-Ligand Binding Site Recognition

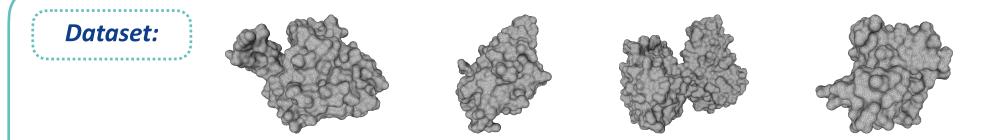
Luca Gagliardi, Andrea Raffo, Ulderico Fugacci, Silvia Biasotti, Walter Rocchia, Hao Huang, Boulbaba Ben Amor, Yi Fang, Yuanyuan Zhang, Xiao Wang, Charles Christoffer, Daisuke Kihara, Apostolos Axenopoulos, Stelios Mylonas, Petros Daras











We extracted protein-ligand complexes from Binding MOAD and processed by:

- + Considering just ligands with significative ligand molecular weight and resolution
- + Removing redundant structures
- **Creating PQR files** using the AMBER force field via the pdb2pqr software
- + Building the triangulation (in OFF format) of the SES molecular surfaces via NanoShaper
- + **Discarding** structures with **multiple connected components**
- + Labeling atoms and triangulation vertices in accordance with the identified binding sites
- + Dropping highly overlapping binding regions

Dataset:

The resulting dataset consists of:

- + 1091 protein structures
- + 1721 binding sites

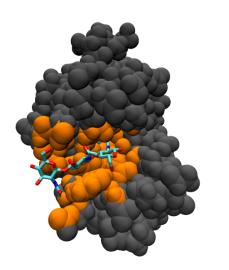
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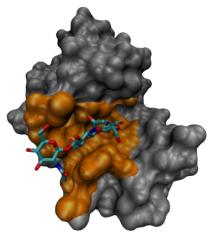
Provided in two different representations:

- + Atom spheres (PQR format)
- + SES surface (OFF format)









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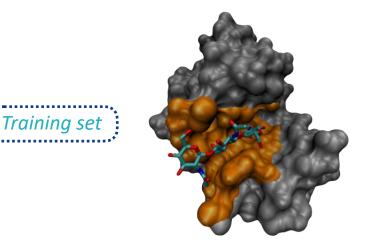
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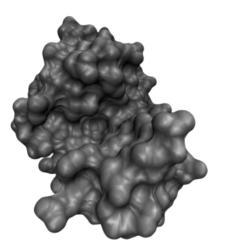
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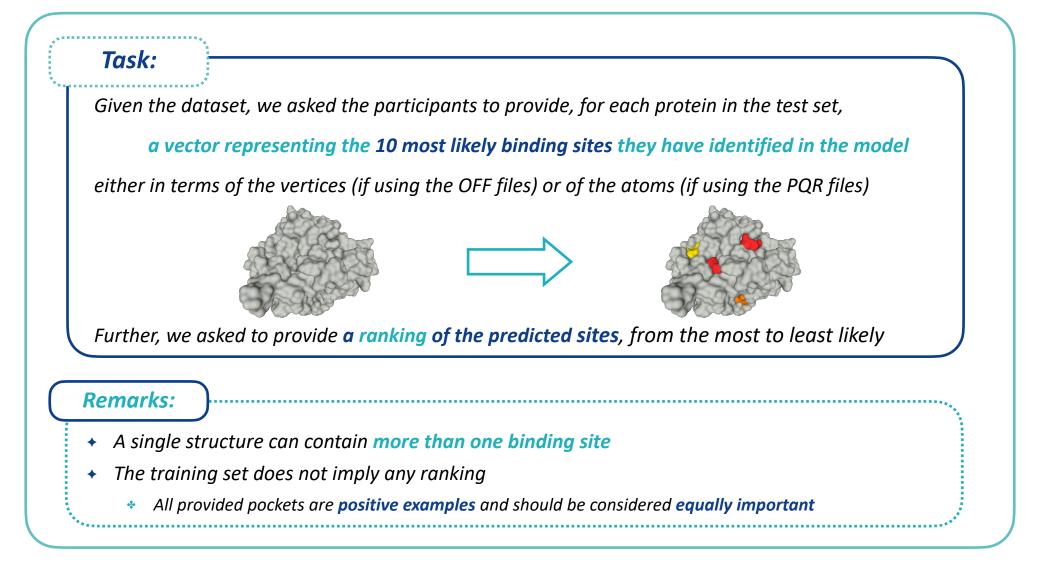
Subdivided into:

- + Training set (85%)
- + (anonymized) Test set (15%)









Methods:

Eight groups from *four different countries* registered to the track *Four* of them proceeded with the *submission* of their results:

- + Method M1 Point Transformer
 - 🔹 by H. Huang, B. Ben Amor, Y. Fang
- + Method M2 GNN-Pocket
 - 🔹 by Y. Zhang, X. Wang, C. Christoffer, D. Kihara
- + Method M3 DeepSurf
 - * by A. Axenopoulos, S. Mylonas, P. Daras
- + Method M4 NS-Volume
 - * by L. Gagliardi, W. Rocchia

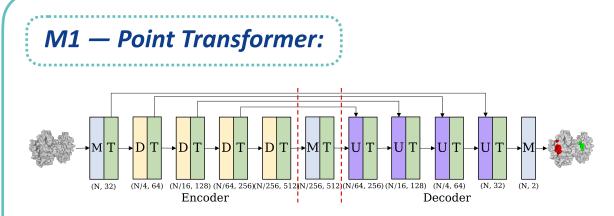
The organizers of the track are L. Gagliardi, A. Raffo, U. Fugacci, S. Biasotti, W. Rocchia



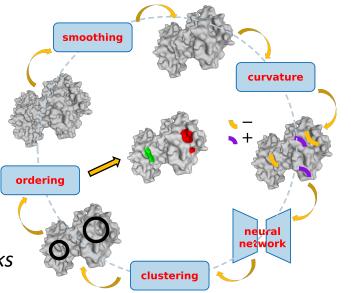
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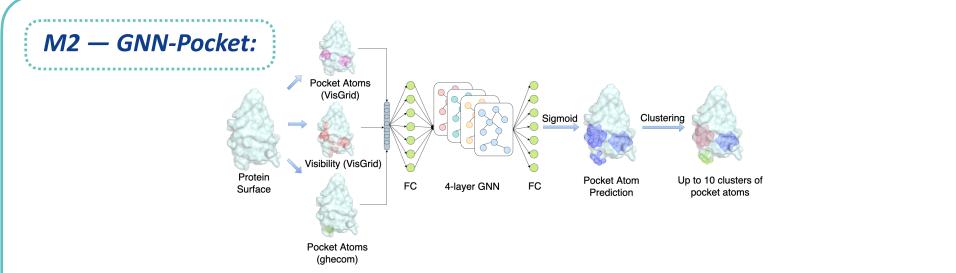


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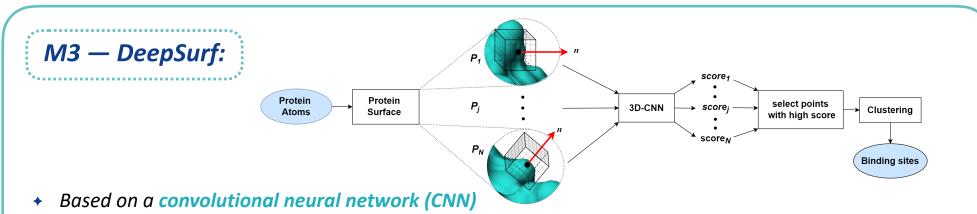


- Originally proposed for machine translation, it has achieved notable performance on various computer vision tasks
- + The neural network model:
 - * Adopts a **U-Net architecture** consisting of an encoder and a decoder
 - Is adapted to learn per-vertex local shape geometric features
 - Is fed with a 5-dimensional vertex feature (coordinates and curvatures) to predict a binary segmentation result as a ligandability score
- Binding regions are obtained by clustering the vertices with a high ligandability score through a density-based algorithm
 - * Regions are *filtered and ranked* on the basis of the *average squared ligandability score* of their vertices

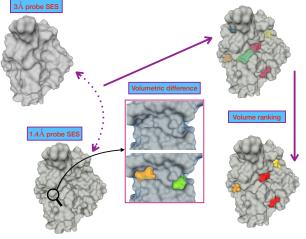




- + Based on a graph neural network (GNN) fed with a feature vector for each atom
- The vector is obtained by concatenating 3 features:
 - * A binary output from VisGrid, which indicates if an atom has a visibility lower than a cutoff
 - * The number of closest grid points that are predicted as pockets by ghecom
 - * The number of grid points within 8 Å that are predicted as pockets by VisGrid
- A bottom-up hierarchical clustering method, which minimizes the distance between the closest pairs of clusters, is adopted to group pocket atoms into pocket regions
 - The top-10 pockets by the sum of probability values of atoms are selected



- A number of local 3D voxelized grids are placed on protein surface and used for extracting features with which feed the network
- + For each protein atom, **18 chemical features** are calculated
 - Each grid voxel receives the features of the atoms inside it
- This requires information on the atom types that lacks in the provided database
 - Such information was **inferred from the atom radii** (regarded as a highly confident indication of the atom type)
- Binding regions are obtained by clustering the vertices with a high ligandability score using the meanshift algorithm
 - * Regions are **sorted** on the basis of the **average ligandability score** of their vertices
- DeepSurf was originally trained on scPDB database
 - 16034 entries corresponding to 4782 proteins with 17594 total binding samples

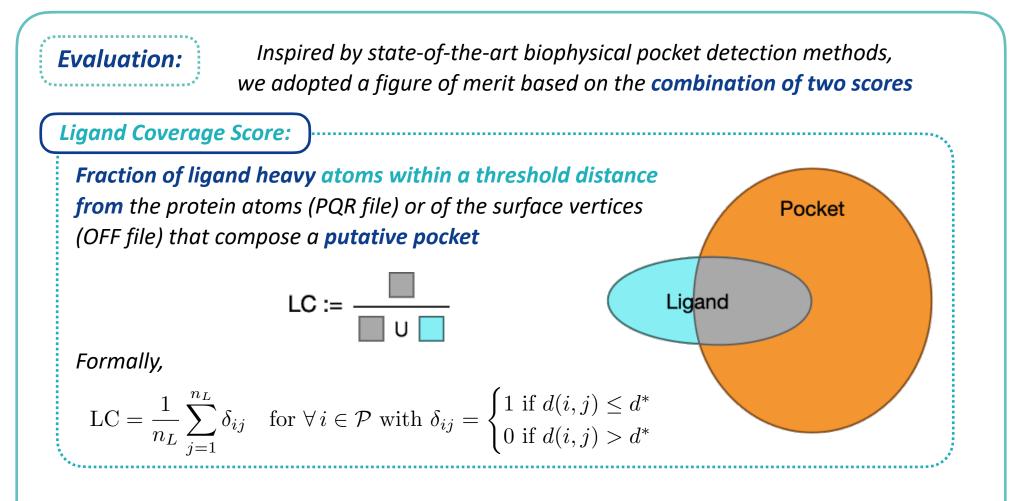


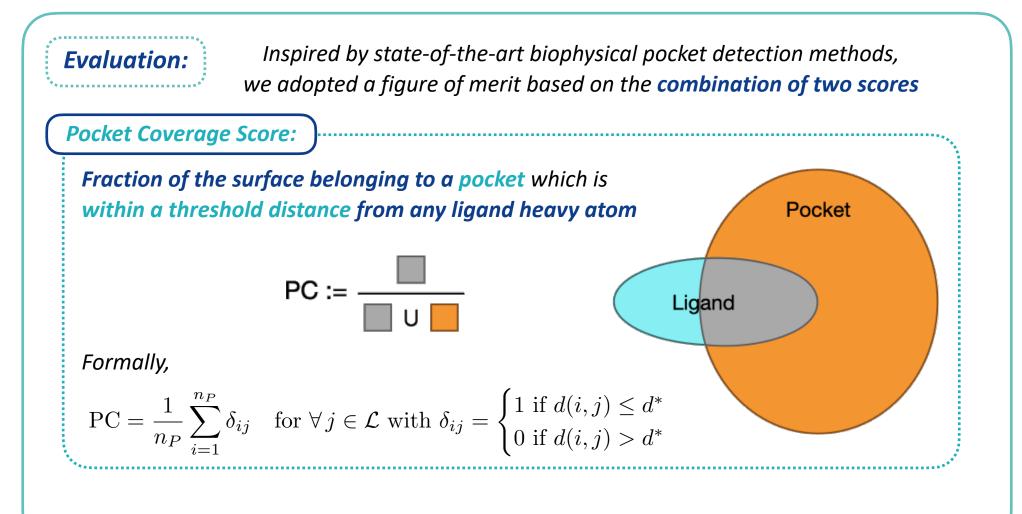
- Based on NanoShaper, an efficient software for the triangulation of molecular surfaces
 - NanoShaper offers also a pocket detection function
- Pockets are defined as the volumetric difference between the space regions enclosed within the SESs of the protein obtained with two different probe radii
 - Points are flagged if **simultaneously** inside the 3 Å SES and outside the 1.4 Å SES (water molecule effective radius)
 - A *filtering procedure* is adopted which preserves points which are
 - (i) within 1.4 Å from all flagged point or
 - (ii) within 1.4 Å from points fulfilling (i)
 - * Pockets are defined as the **unconnected components** after the filtering by applying a **flood-fill procedure**
- Obtained pockets are sorted by volume

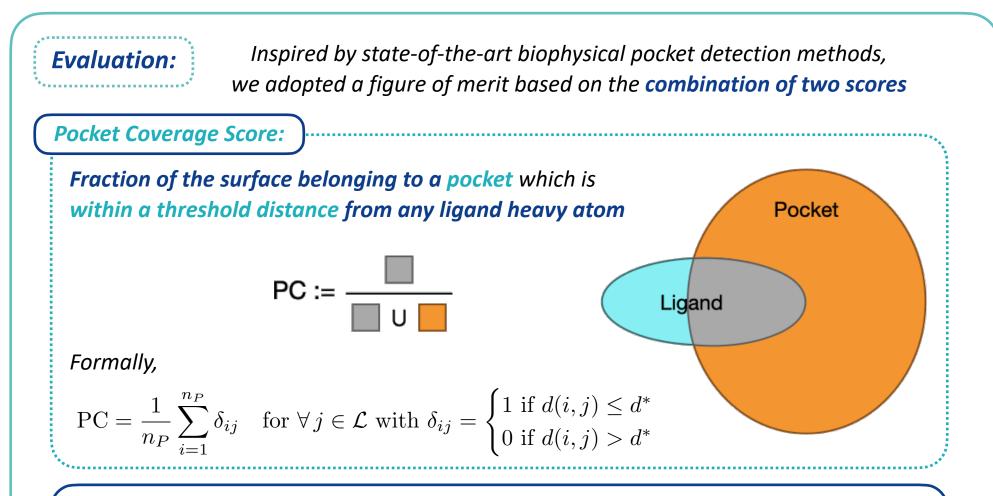
M4 – NS-Volume:

Evaluation:

Inspired by state-of-the-art biophysical pocket detection methods, we adopted a figure of merit based on the **combination of two scores**







A putative pocket is correctly matched if it scores at least 50% in LC and at least 20% in PC

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Comparison:	Method	Top1	Top3	Top10	LC	PC	nPockets
***	M1 - Point Transformer	69.1	75.9	75.9	96.4	60.4	2.1
	M2 - GNN-Pocket	53.4	54.6	55.4	93.7	47.5	1.9
	M3 - DeepSurf	87.6	89.2	89.2	95.0	67.9	1.6
	M4 - NS-Volume	59.0	76.7	83.9	88.8	74.8	11.6
♦ We report	Fpocket	60.2	75.1	84.7	92.5	64.7	8.9

- * Average ranking in terms of **Top1**, **Top3**, and **Top10** performance
- Average LC and PC scores over successfully predicted pockets
- * Average number of generated pockets per structure
- Results are expressed as the percentage of success rate normalized over the total number of structure-ligand pairs
- For sake of comparison, we report also the results obtained by Fpocket on the same dataset
 - * A standard and **well established tool** for pocket detection
 - Not eligible as a competing method in the SHREC track since it considers also chemical features and not just geometrical ones

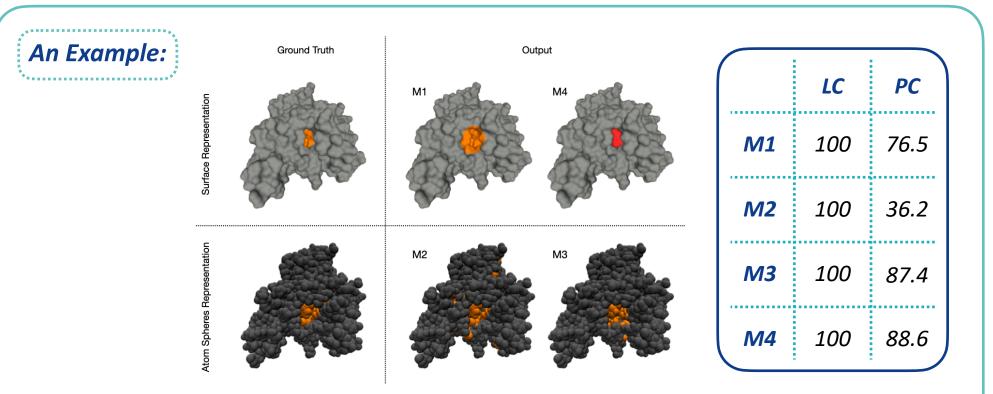
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+ M3 shows an excellent performance

- * Obtained results outstand also Fpocket
- Despite the small number of putative pockets generated, these are extremely well predicted
- * Information leveraged goes **beyond pure geometry** and the training set is larger than the one provided
- **On Top10**, M4 and Fpocket obtain similar scores to M3
- + Only M4 and Fpocket return more than about 2 putative pockets per structure on average
- + All methods perform very well in term of Ligand Coverage score
- A significantly lower Pocket Coverage score is measured

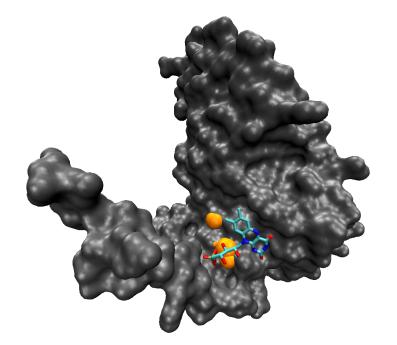


 Low average PC score indicates that a method is prone to generate pockets which are larger than the binding ligand

 We observe that, in general, M2 generates pockets which are often larger than the binding region and scattered into disconnected segments

Discussion:

Proposed methods have **difficulties** in identifying particularly **shallow binding sites**



This is due to:

- Methods completely relying on geometry for the generation of putative pockets are optimized to recognize cleft and cavities (which are often found to contain binding ligands)
- Shallow pockets attain the role of binding site mainly for their chemical properties rather than their shape
- Shallow sites are rare in the training set

Conclusions:

- We proposed a SHREC track aimed at evaluating the effectiveness of computational methods based purely on geometrical information in the detection of binding sites on protein surfaces
- We created a dataset of protein structures expressed both in terms of atom spheres and SES surface representations and we enriched the training set with positive examples of known ligandable pockets
- We analyzed and compared four different proposed methods on the basis of two evaluation measures:
 - Most of the proposed methods show very good performance
 - All methods struggle in the recognition of shallow binding sites
 - Proposed methods generally perform *low Pocket Coverage* score
- Future directions and possible improvements:
 - Problem in this SHREC track is an instance of a **one-class discrimination task**
 - Low PC scores suggests the possibility of considering a higher segmentation of the returned sites into separate smaller pockets or sub-units

Resources:

- Dataset, benchmark, predictions of participants that originated the results: <u>https://github.com/concept-lab/shrec22_proteinLigandBenchmark</u>
- + M1 Point Transformer:

https://github.com/aaron-h-code/Protein_SHREC2022/

+ M2 – GNN-Pocket:

https://github.com/kiharalab/GNN_pocket

+ M3 – DeepSurf:

https://github.com/stemylonas/DeepSurf_SHREC22

+ M4 – NS-Volume:

https://github.com/concept-lab/NS_pocket

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Thank you for the attention!

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